

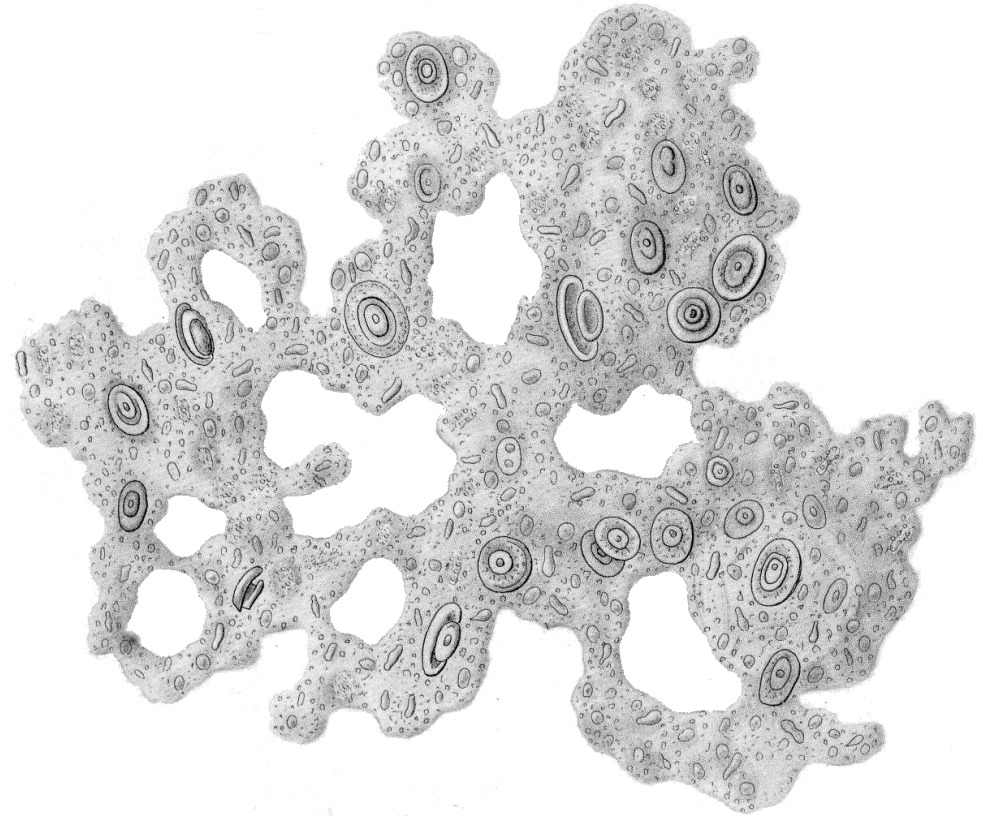
Devolution

*Darwin's Mechanism Works
Chiefly by Squandering Genetic
Information for Short-term Gain*

Michael J Behe
Lehigh University
Bethlehem, PA

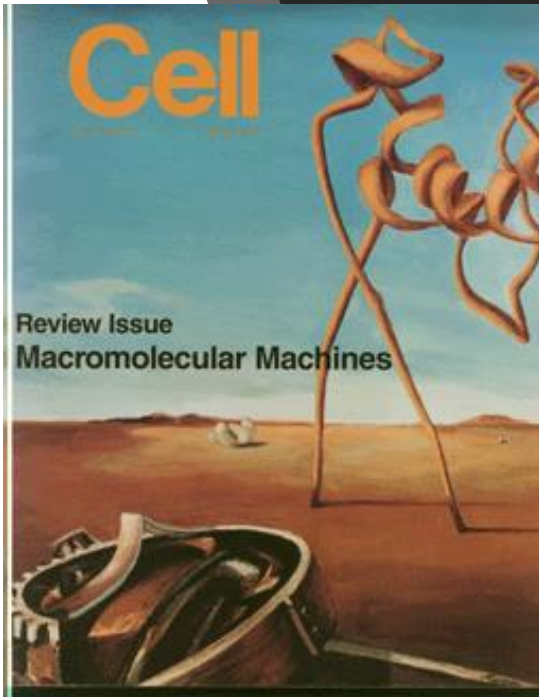
Bathybius haeckelii
1870

“Protoplasm”



Cell (1998) 92, table of contents.

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- **Polymerases and the Replisome: **Machines** within **Machines****, Tania A Baker and Stephen P Bell
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- **Molecular Movement inside the Translational **Engine****, Kevin S Wilson and Harry F Noller
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Cryo-EM structure of the entire mammalian F₁-type ATP synthase

Gergely Pinke, Long Zhou and Leonid A. Sazanov

The majority of adenosine triphosphate (ATP) powering cellular processes in eukaryotes is produced by the mitochondrial F₁F₀ ATP synthase. Here, we present the atomic models of the membrane F₀ domain and the entire mammalian (ovine) F₁F₀, determined by cryo-electron microscopy. Subunits in the membrane domain are arranged in the 'proton translocation cluster' attached to the c-ring and a more distant 'hook apparatus' holding subunit e. Unexpectedly, this subunit is anchored to a lipid 'plug' capping the c-ring. We present a detailed proton translocation pathway in mammalian F₀ and key inter-monomer contacts in F₁F₀ multimers. Cryo-EM maps of F₁F₀ exposed to calcium reveal a retracted subunit e and a disassembled c-ring, suggesting permeability transition pore opening. We propose a model for the permeability transition pore opening, whereby subunit e pulls the lipid plug out of the c-ring. Our structure will allow the design of drugs for many emerging applications in medicine.

The ATP synthase (F₁F₀) employs a unique rotary mechanism, harvesting the proton motive force (PMF) created during respiration in mitochondria by electron transport chain (ETC) complexes^{1,2}. The ATP synthase/ATPase family comprises membrane-bound protein complexes responsible either for ATP synthesis, utilizing PMF (F-type and A-type), or for establishing PMF using the energy released from ATP hydrolysis (V-type)^{3,4}. F-type enzymes produce ATP in bacteria, chloroplasts and mitochondria, while V-ATPases (vacuolar) acidify the interior of eukaryotic intracellular compartments. The F₁F₀ complex consists of a soluble F₁ domain, responsible for the synthesis of ATP, and a membrane F₀ domain, involved in proton translocation. These domains are connected by a central stalk rotating inside the F₁ and a stationary peripheral stalk (PS)^{5,6}. During ATP synthesis, PMF-driven rotation of the c-ring in F₀ is transmitted via the central stalk to power the conformational changes in the F₁, resulting in the synthesis of one ATP molecule per 120° rotation (because F₁ is three-fold symmetric).

F₁F₀ plays other important roles apart from energy generation. ETC complexes I–IV are mostly organized into supercomplexes^{7–9} in flat regions of the inner mitochondrial membrane (IMM)¹⁰. F₁F₀, on the other hand, forms rows of dimers along the highly curved cristae ridges, thus shaping them¹¹. The enzyme is also implicated in the formation of the permeability transition pore (PTP), which triggers cell death^{12,13}.

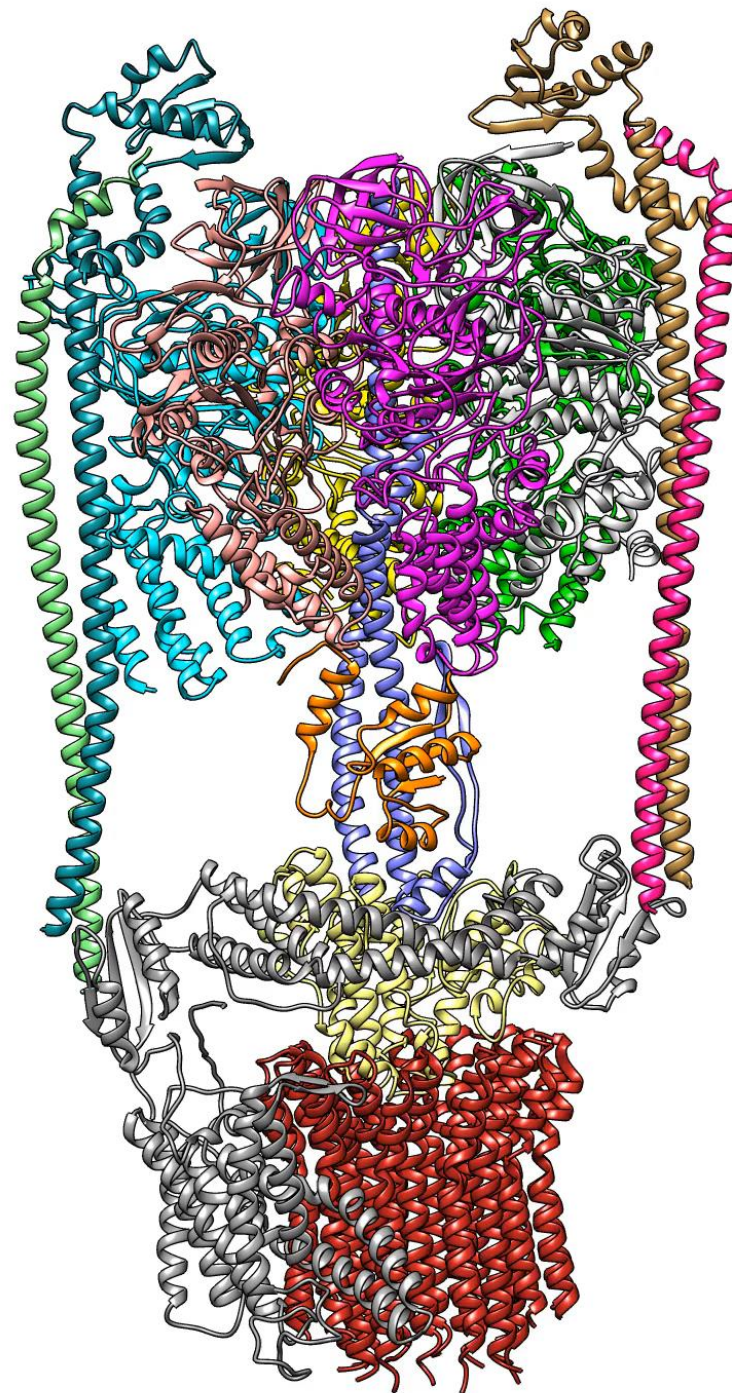
PTP opening can be triggered by the accumulation of Ca²⁺ or by intense oxidative stress, characterizing ischemia-reperfusion injury^{14,15}. The initial opening of the PTP is reversible, establishing a 2–3-nm pore, followed by mitochondria swelling and rupture, the release of pro-apoptotic factors such as cytochrome c and cell death^{16,17}. The molecular nature of the PTP is controversial. The mitochondrial matrix protein cyclophilin D (CyPD)¹⁸ sensitizes the PTP to Ca²⁺. CyPD binding to its partners is blocked by cyclosporin A (CsA), which inhibits the PTP¹⁹. The recent discovery that CyPD binds to F₁F₀ subunit OSCP opened up the possibility that F₁F₀ forms the PTP²⁰. Many recent studies have both supported^{21–23} and refuted^{24–28} the still hotly debated role of F₁F₀ in the PTP (Supplementary Note 1). Several mutagenesis studies converge on the c-ring as a possible location of the pore^{18,29,30}.

We have previously determined the first atomic structure of V/A-ATPase as a representative of the V-type family³¹. Structures of entire bacterial³², yeast³³ and chloroplast³⁴ F-type ATP synthases have also been determined recently. However, knowledge about the arguably most important representative of the family—mammalian mitochondrial ATP synthase—remains incomplete. Crystallography has revealed many structures of F₁ subcomplexes^{35,36}, as have cryo-EM studies on the entire complex³⁷. The recent porcine enzyme model is the most complete so far³⁷. However, due to the limited resolution in the membrane domain, four subunits were modeled as poly-alanine and three more were completely misplaced, so the atomic model for most of the membrane domain remains unknown.

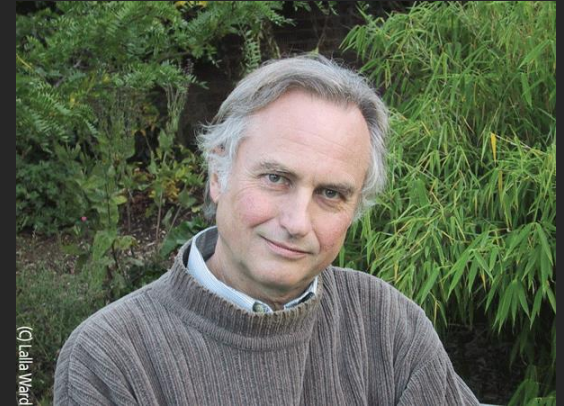
Detailed knowledge about the F₀ domain is of crucial importance because this is where the proton translocation takes place and where the monomers interact to form physiological dimers. Here, we address these questions by solving the structure of the entire mammalian F₁F₀.

Results

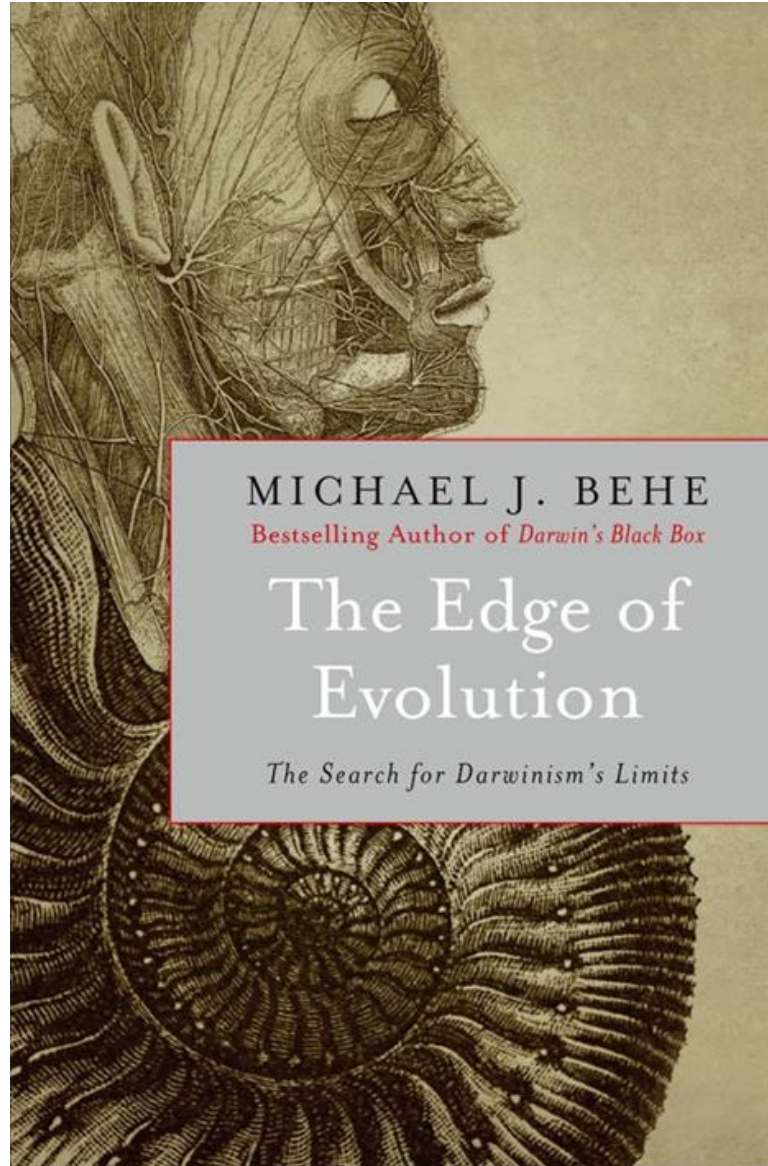
Structure determination. We purified ATP synthase from ovine heart mitochondria in the mild detergent laurylmaltose neopentylglycol (LMNG) and collected two datasets, from the 'monomer' and 'multimer' fractions (Extended Data Fig. 1a–c). The most populated and best resolved ground state of the monomer (Extended Data Fig. 1d) is similar to the previously observed (at lower resolution) state 1a of bovine enzyme (PDB 5ARA)³⁸. The other two main rotational states (resulting from ~120° rotation of the central stalk subunit γ) were only at ~7–8-Å resolution due to the lower number of particles (Extended Data Fig. 2). Further 'in-between' states were also present, but with some of the α/β subunits disordered, possibly due to lower enzyme stability in such states. State-1a F₁F₀ maps were refined to 3.8-Å resolution overall (Extended Data Figs. 1d and 3d), with focused refinements reaching 3.5 Å for the F₁ domain and 4.2 Å for F₀ (obtained using a novel strategy of weighted masks; Methods). Focusing on F₀ classification of particles in all rotational states revealed that the majority of particles classify into one consensus class, producing, after Fo-focused refinement, a 3.8-Å-resolution map (Extended Data Fig. 3e). This map was well resolved at the side chain level in all Fo areas (Extended Data Fig. 4e), suggesting that,



Dawkins R. 1986. *The Blind Watchmaker*. New York: Norton, p. 43



“We have seen that living things are too improbable and too beautifully ‘designed’ to have come into existence by chance. How, then, did they come into existence? ... **by gradual, step-by-step transformations from simple beginnings...**”

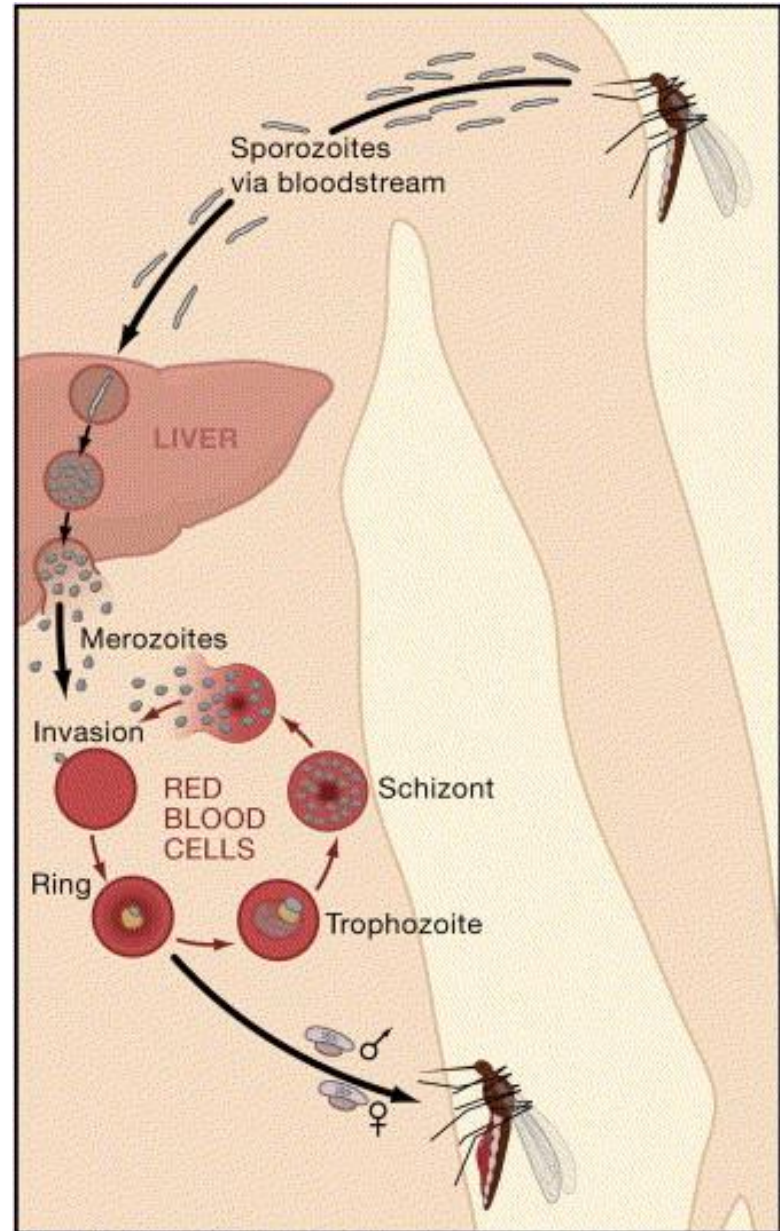


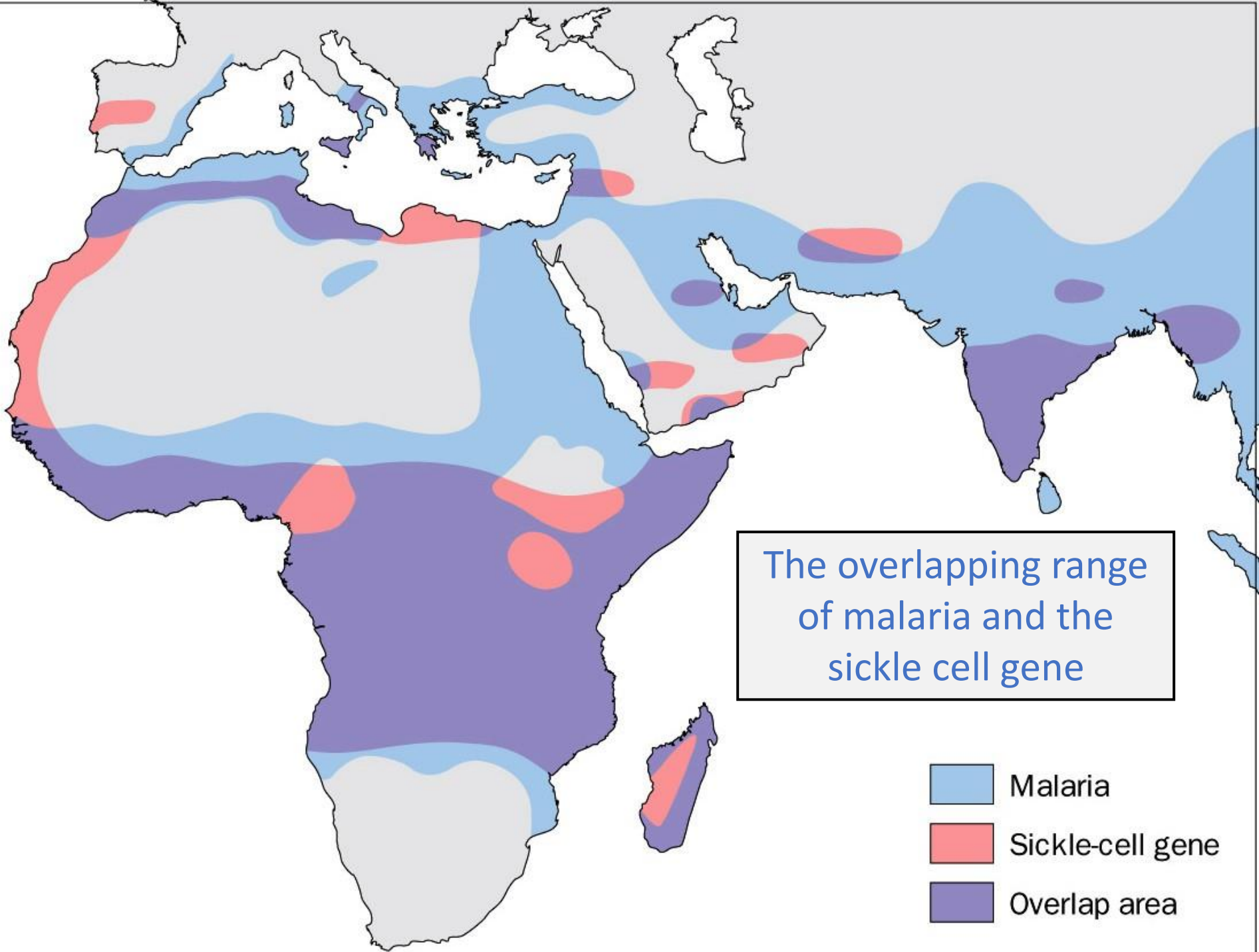
2007

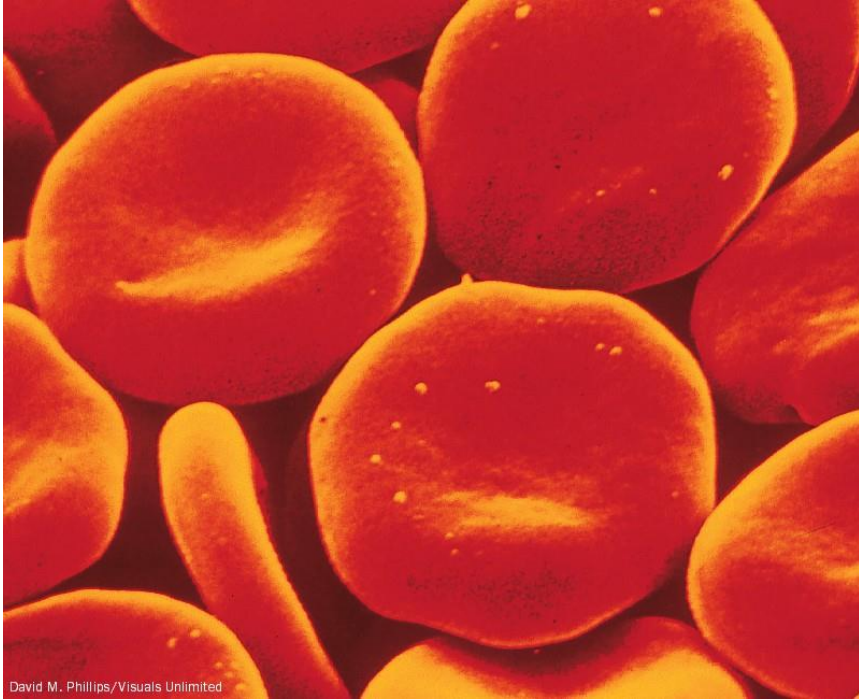
What is our
best evidence
of what
Darwinian
processes can
actually do?

- The best evidence we have to assess the abilities of Darwinian processes comes from studies of *malaria*, both in genetic changes of humans and in the parasite (*Plasmodium falciparum*) itself.
- Reasons:
 - Detailed genetic studies
 - Sheer population sizes

Infection of a human by the malarial parasite, *Plasmodium falciparum*







David M. Phillips/Visuals Unlimited

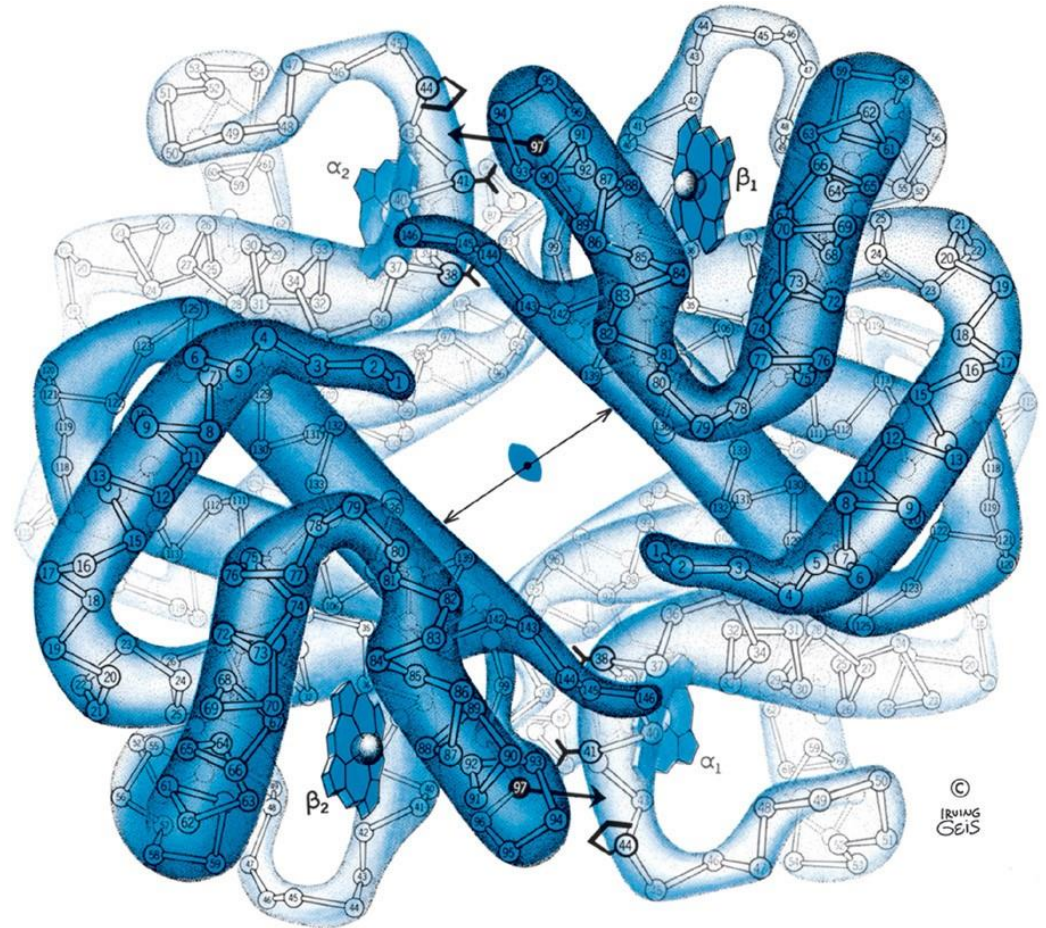


Bill Longcore/Photo Researchers

Normal red blood cells

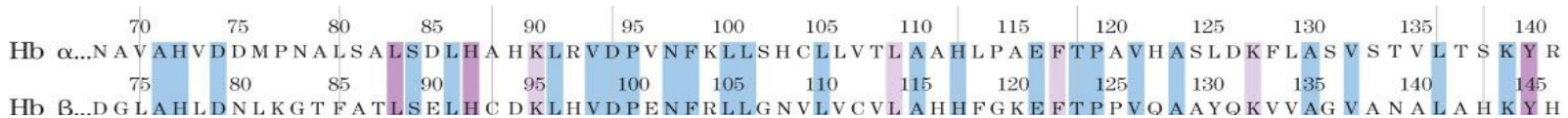
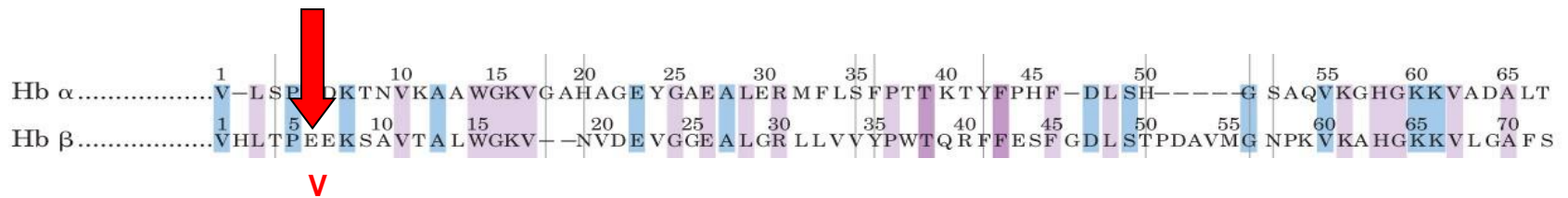
Sickle red blood cells

The molecular structure of hemoglobin

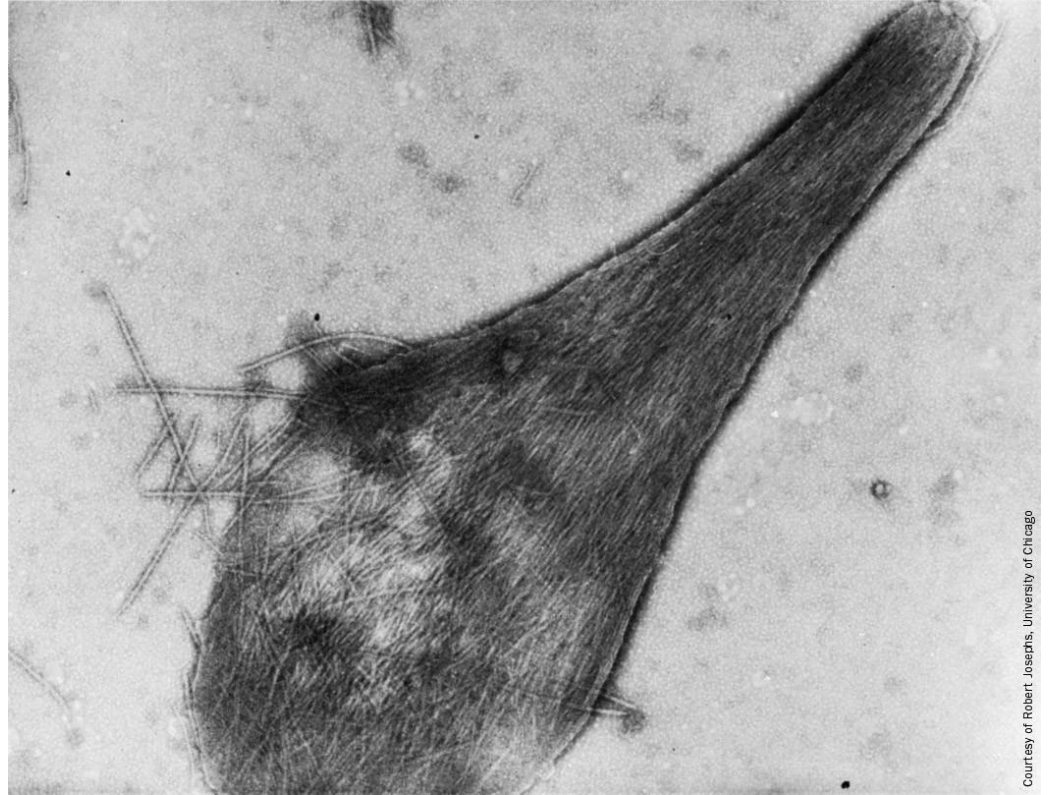


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The Amino Acid Sequences of the α and β Chains of Human Hemoglobin



Electron
micrograph of
deoxygenated
sickle hemo-
globin fibers



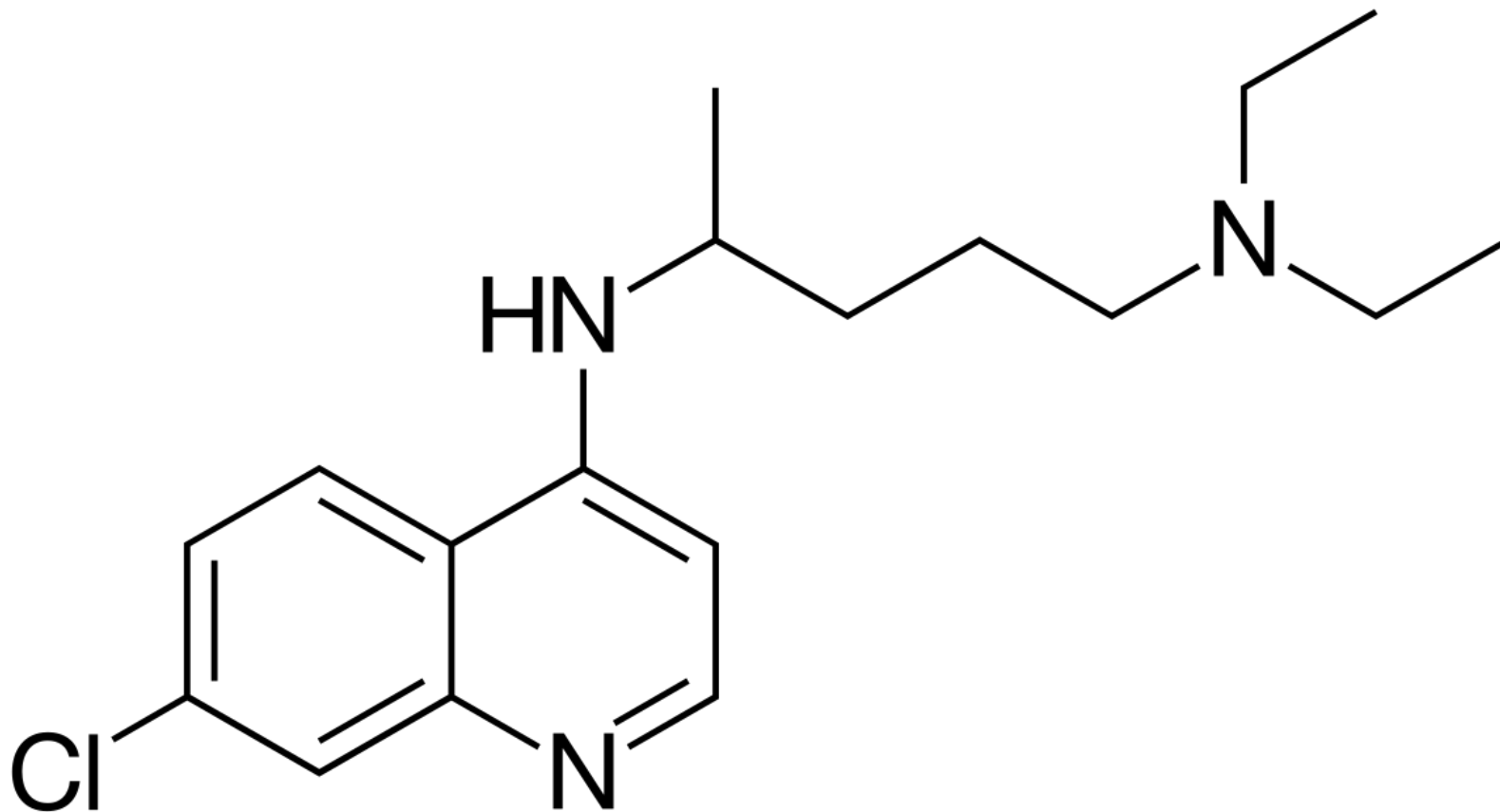
Human genetic effects selected for resistance to malaria

<u>Gene</u>	<u>Mutation</u>	<u>Adverse effects</u>
Hemoglobin	HbS	Sickle cell disease
	alpha-thalassemia	Anemia/ broken gene
	beta-thalassemia	Anemia/ broken gene
	Hereditary persistence of fetal hemoglobin	Broken genetic controls
G6PD	Point mutations, deletions	Anemia / decrease or loss of G6PD function
Band 3 protein	deletion	Lethal in two copies / broken gene
Duffy antigen	Point mutation	Protein expression lost in red blood cells

*Plasmodium
falciparum*



Chloroquine



A Requiem for Chloroquine

I. M. Hastings, P. G. Bray, S. A. Ward

Chloroquine (CQ) has historically been the mainstay of malaria treatment, particularly in the worst affected regions of sub-Saharan Africa. The recent development of widespread CQ resistance in *Plasmodium falciparum*, the most dangerous of the four malaria parasite species, has contributed significantly to escalating mortality rates in Africa (1) and to the resurgence of malaria as an immediate public health priority (2). Several pressing scientific questions have emerged within the context of this humanitarian disaster: What is the molecular basis for CQ resistance, and how has this influenced the dynamics of resistance? Why did CQ remain effective for 20 years, yet

its immediate replacement sulfadoxine-pyrimethamine (SP) last less than 5 years? Has the widespread deployment of CQ jeopardized the use of other drugs targeting the same parasite biochemical pathways? As reported on page 210 of this issue, Sidhu *et al.* (3) have obtained data relevant to all three questions by creatively exploiting the *pfert* gene, which encodes a putative transporter protein in the digestive vacuole membrane of the malaria parasite. They replaced the endogenous *pfert* gene in a CQ-sensitive strain of *P. falciparum* with a *pfert* gene from each of three CQ-resistant strains. All such replacement strains ("constructs") showed CQ resistance in vitro, demonstrating that *pfert* mutations are sufficient, within their selected genetic background, to encode resistance. Reduced levels of *pfert* gene expression in the constructs also showed that up-regulation of *pfert* is not required for resistance. Next,

the authors investigated cross-resistance between CQ and other antimalarial drugs.

Previous work from this and other groups has implicated eight or nine different mutations in the development of CQ resistance (4). The sequential accumulation of these mutations plausibly explains the observed genetics and epidemiology of CQ resistance (see the figure). So why did CQ persist so much longer than SP as a frontline antimalarial? First, four sequential mutations in the *dhfr* gene—which encodes dihydrofolate reductase, an enzyme essential for pyrimethamine—appear sufficient for SP resistance (5). These four mutations accumulate much faster than the nine required for CQ resistance. Second, CQ persists at therapeutically useful concentrations for a shorter period than SP, leading to lower selection pressures for resistance (6). Third, resistance may involve genes other than *pfert*, such that sexual recombination during the malaria life cycle breaks down genetic combinations, slowing resistance (7, 8). The relative involvement of other genes remains controversial. Sidhu *et al.* show that *pfert* is

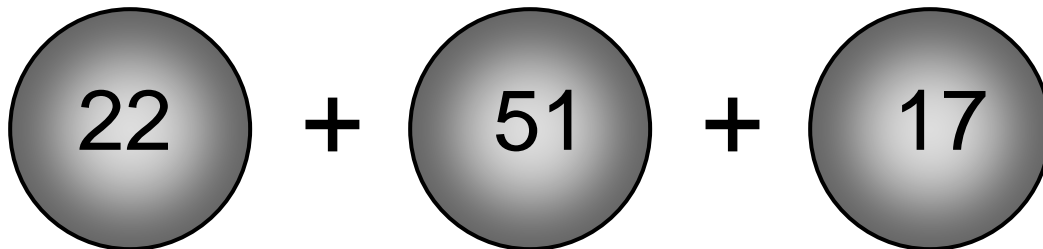
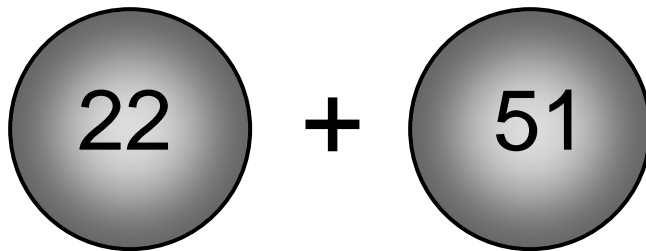
The authors are at the Liverpool School of Tropical Medicine, Pembroke Place, Liverpool L3 5QA, UK. E-mail: hastings@liverpool.ac.uk; p.g.bray@liverpool.ac.uk; saward@liverpool.ac.uk



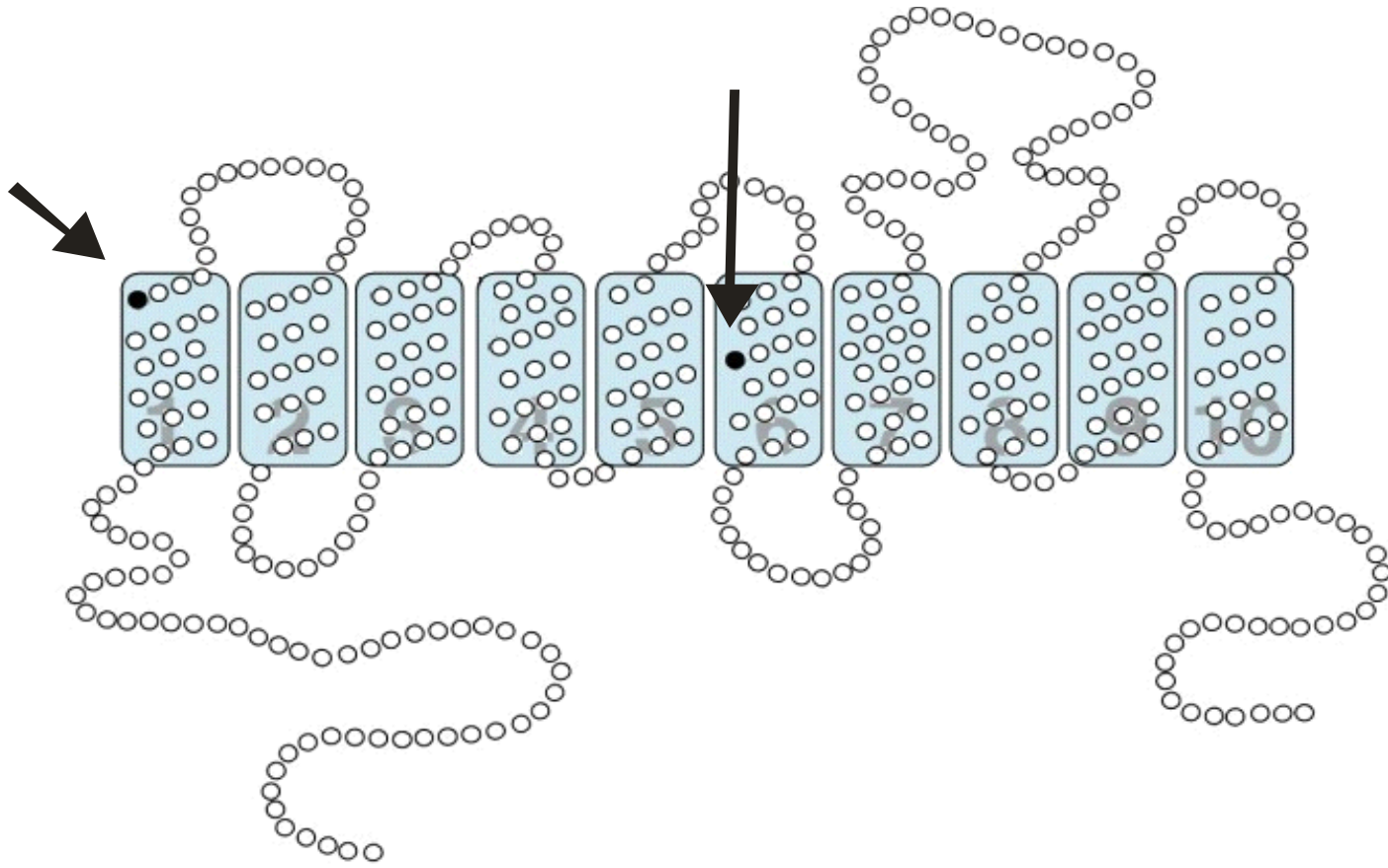
Frequency of the
development of
antibiotic resistance
of *P. falciparum*

- Resistance to atovaquone arises in about every **third** patient (about 1 in 10^{12} cells)
 - (Looareesuwan, S., et al. 1996. Clinical studies of atovaquone, alone or in combination with other antimalarial drugs, for treatment of acute uncomplicated malaria in Thailand. *Am. J. Trop. Med. Hyg.* **54**:62-66)
- Resistance to chloroquine arises in about every **billionth** patient (about 1 in 10^{20} cells)
 - (White NJ. 2004. Antimalarial drug resistance. *J Clin Invest* **113**:1084-1092.)

Lottery: The difficulty of matching several numbers



Chloroquine-resistance in malaria requires several mutations



This is not an
argument that
Darwinism *cannot*
make complex
functional systems;
it is an *observation*
that it *does not*.

No unintelligent process helped much with malarial chloroquine-resistance, including:

- Darwinism
- Self-organization
- Self-engineering
- Symbiosis
- *Nor any as-yet-undiscovered process*



DARWIN *Devolves*

The New Science About DNA
That Challenges Evolution



MICHAEL J. BEHE

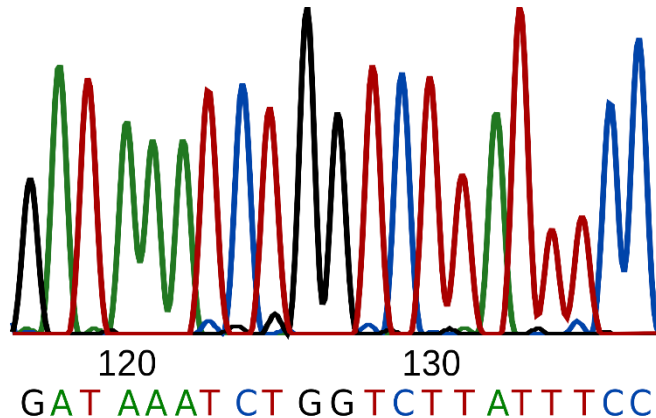
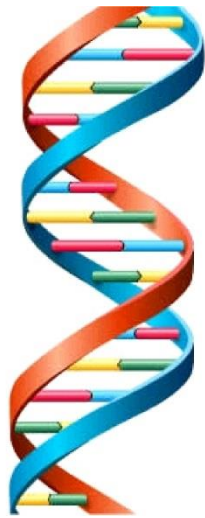
Author of *DARWIN'S BLACK BOX*

2019



Science
advances
with new
technology

The DNA Sequencing Revolution



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170 180 190  
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GATGAAGAACGCAGCGAAACCGGATATGTAAT
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**Key
Concepts
from
*Darwin
Devolves***

The First Rule of
Adaptive Evolution

The Principle of
Comparative Difficulty

The Family Line

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Richard Lenski

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Richard E. Lenski



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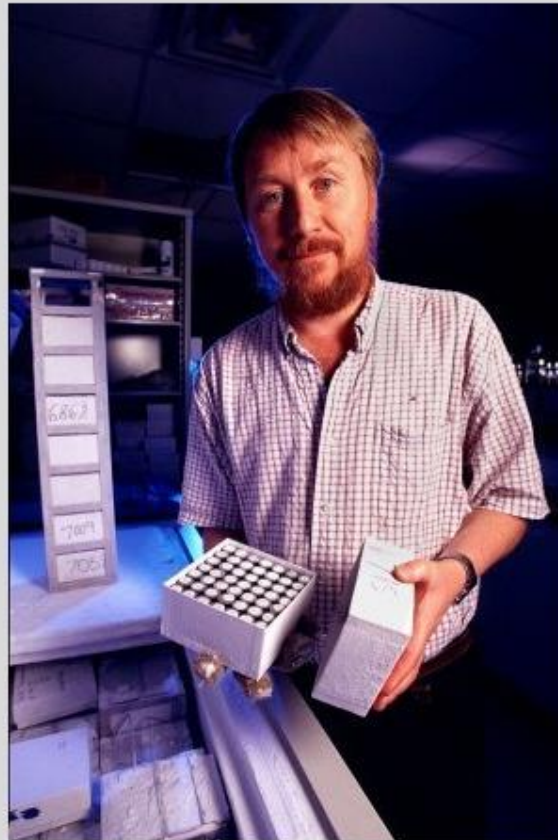


Photo courtesy of Bruce Fox, MSU

Richard E. Lenski
Hannah Distinguished
Professor
Michigan State
University

Email: lenski@msu.edu

Last Updated: 7 April 2007

Feature article in [Science](#) magazine--"[Test tube evolution catches time in a bottle](#)" by Tim Appenzeller--on our research with evolving bacteria

[Column on our research with digital organisms](#) from [Natural History](#) magazine [Posted with permission of author [Carl Zimmer](#), illustrator [James Marsh](#), and [Natural History](#) magazine.]

Also of interest ...

- ♦ [The *E. coli* long-term evolution experiment](#)
- ♦ [Some thoughts and readings on the history and philosophy of science](#)

“expression of both the ribose operon and the maltose regulon *decreased* after 20,000 generations of experimental evolution. These changes may therefore reflect *beneficial* mutations in these regulons. Indeed, *deletions* of the *rbs* operon were found previously in all 12 of the evolved populations.”

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DOI: 10.1534/genetics.105.049619

Parallel Changes in Global Protein Profiles During Long-Term Experimental Evolution in *Escherichia coli*

Ludovic Pelosi,* Lauriane Kühn,[†] Dorian Guetta,* Jérôme Garin,[†] Johannes Geiselmann,*
Richard E. Lenski[‡] and Dominique Schneider*^{*,1}

*Laboratoire Adaptation et Pathogénie des Microorganismes, Université Joseph Fourier, CNRS UMR 5163, 38041 Grenoble, France,
[†]Laboratoire Chimie des Protéines DRDC-CP, ERM 0201 CEA/INSERM/UJF, 38054 Grenoble, France and [‡]Department of
Microbiology and Molecular Genetics, Michigan State University, East Lansing, Michigan 48824

Manuscript received August 15, 2005

Accepted for publication May 11, 2006

ABSTRACT

Twelve populations of *Escherichia coli* evolved in and adapted to a glucose-limited environment from a common ancestor. We used two-dimensional protein electrophoresis to compare two evolved clones, isolated from independently derived populations after 20,000 generations. Exceptional parallelism was detected. We compared the observed changes in protein expression profiles with previously characterized global transcription profiles of the same clones; this is the first time such a comparison has been made in an evolutionary context where these changes are often quite subtle. The two methodologies exhibited some remarkable similarities that highlighted two different levels of parallel regulatory changes that were beneficial during the evolution experiment. First, at the higher level, both methods revealed extensive parallel changes in the same global regulatory network, reflecting the involvement of beneficial mutations in genes that control the ppGpp regulon. Second, both methods detected expression changes of identical gene sets that reflected parallel changes at a lower level of gene regulation. The protein profiles led to the discovery of beneficial mutations affecting the *malT* gene, with strong genetic parallelism across independently evolved populations. Functional and evolutionary analyses of these mutations revealed parallel phenotypic decreases in the maltose regulon expression and a high level of polymorphism at this locus in the evolved populations.

EXPERIMENTAL EVOLUTION, LOSS-OF-FUNCTION MUTATIONS, AND "THE FIRST RULE OF ADAPTIVE EVOLUTION"

MICHAEL J. BEHE

Department of Biological Sciences, Lehigh University, Bethlehem, Pennsylvania 18015 USA

E-MAIL: MJB1@LEHIGH.EDU

KEYWORDS

experimental evolution, adaptation, mutation, loss of function, malaria,
Yersinia pestis

ABSTRACT

Adaptive evolution can cause a species to gain, lose, or modify a function; therefore, it is of basic interest to determine whether any of these modes dominates the evolutionary process under particular circumstances. Because mutation occurs at the molecular level, it is necessary to examine the molecular changes produced by the underlying mutation in order to assess whether a given adaptation is best considered as a gain, loss, or modification of function. Although that was once impossible, the advance of molecular biology in the past half century has made it feasible. In this paper, I review molecular changes underlying some adaptations, with a particular emphasis on evolutionary experiments with microbes conducted over the past four decades. I show that by far the most common adaptive changes seen in those examples are due to the loss

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MICHAEL J. BEHE

Department of Biological Sciences, Lehigh University, Bethlehem, Pennsylvania 18027 USA

E-MAIL: MJB1@LEHIGH.EDU

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Adaptive evolution can cause a species to gain, lose, or modify a function; therefore, it is of basic interest to determine whether any of these events dominates the evolutionary process under particular circumstances. Because mutations occur at the molecular level, it is necessary to measure the molecular changes produced by the underlying processes in order to assess whether a given adaptation is best considered as a gain, loss, or modification of function. Although that was once impossible, the advent of molecular biology in the past half century has made it possible. In this paper, I review molecular changes underlying some adaptations, with a particular emphasis on evolutionary experiments with microbes conducted over the past few decades. I show that for the most common adaptive changes seen in those examples are due to the loss or modification of a pre-existing molecular function, and I discuss the possible reasons for the preponderance of such mutations.

ADAPTATION BY GAIN, LOSS, OR
MODIFICATION OF FUNCTION

IN *THE ORIGIN OF SPECIES*, Darwin (1859) emphasized the rolelessness of natural selection:

[N]atural selection is daily and hourly scrutinizing, throughout the world, every variation, even the slightest, rejecting that which is bad, preserving and adding up all that is good, silently and inensibly working, whenever and wherever opportunity offers, at the improvement of each organic being in relation to its organic and inorganic conditions of life. (p. 94)

Yet he realized that the changes that were selected to adapt an organism to its environment did not have to be ones that conferred upon it a new ability, such as flight or flight. For instance, while observing some barnacles, Darwin discovered unexpected cases of the gross simplification of an organism (Stott 2005):

The male is as transparent as glass. . . . In the lower part we have an eye, & great teeth, & vessels seminal in the caputulum we have nothing but a tremendous long penis coiled up & which can be exerted. There is no mouth or stomach nor (1843, vol. 1, p. 104)

The Quarterly Review of Biology, December 2010, Vol. 85, No. 4
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0033-5770/2010/85040023\$12.00

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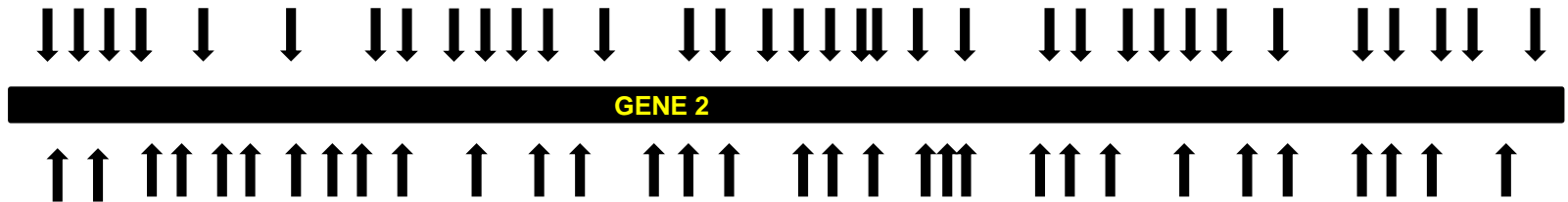
Behe, M. J., 2010 Experimental Evolution, Loss-of-function Mutations, and “The First Rule of Adaptive Evolution”. *Quarterly Review of Biology* 85: 1-27.

The First Rule of Adaptive Evolution:
Break or **blunt** any functional coded element whose loss would yield a net fitness gain.

Points where a gene might be improved



Points where a gene might be broken





Helpful degradation

What observation demonstrates about random mutations

Of those mutations that affect an organism, about 99% are detrimental

Of even *beneficial* mutations, the great majority break genes or degrade function

Darwin's mechanism:

- is dominated by “**Poison-Pill**” mutations: positively-selected, loss-of-function mutations
- **squanders** genetic information for short-term gain



Variation in *Canis lupus familiaris*



Dog breed mutations

phenotype	gene	mutation	ref.
coat color, yellow	Mc1r (melanocortin 1 receptor)	LOF	1
“ , black	CBD103 (β -defensin)	Δ G23	1
coat variation (“furnishings”, hair length, curl)	RSPO2 (R-spondin–2) FGF5 (fibroblast growth factor–5) KRT71 (keratin-71)	167 bp ins UTR C95→F R151→W	2
size	IGF1 (insulin-like growth factor 1)	synonymous SNP	3
short legs	FGF4 (fibroblast growth factor–4)	retroinsertion	4
short muzzle	THBS2 (thrombospondin) SMOC2	multiple SNPs “	5
muscle mass	MSTN (myostatin)	premature stop	6
white spotting	MITF (microphthalmia-assoc. TF)	SINE in reg region	7
hair ridge	FGF3,4,19 (fibroblast gfs)	133-kb duplication	8

[1] Candille,S.I. et al. 2007. A β -defensin mutation causes black coat color in domestic dogs. Science 318:1418-1423.

[2] Cadieu,E. et al. 2009. Coat variation in the domestic dog is governed by variants in three genes. Science 326:150-153.

[3] Sutter,N.B. et al. 2007. A single IGF1 allele is a major determinant of small size in dogs. Science 316:112-115.

[4] Parker,H.G. et al. 2009. An expressed fgf4 retrogene is associated with breed-defining chondrodysplasia in domestic dogs. Science 325:995-998.

[5] Bannasch,D. et al. 2010. Localization of canine brachycephaly using an across breed mapping approach. PLoS. One. 5:e9632.

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[7] Karlsson,E.K. et al. 2007. Efficient mapping of mendelian traits in dogs through genome-wide association. Nat. Genet. 39:1321-1328.

[8] Salmon Hillbertz,N.H. et al. 2007. Duplication causes hair ridge and predisposition to dermoid sinus in Ridgeback dogs. Nat. Genet. 39:1318-1320.



Polar bear evolution

Li, S. et al. 2014.
Population Genomics
Reveal Recent Speciation
and Rapid Evolutionary
Adaptation in Polar Bears.
Cell **157**:785-794.

“[W]e assessed the
impact of polar-bear-
specific substitutions
... by computational
predictions: *a large
proportion (ca. 50%)
of mutations were
predicted to be func-
tionally damaging.*”



Population Genomics Reveal Recent Speciation and Rapid Evolutionary Adaptation in Polar Bears

Shiping Liu,^{1,2,3,4} Elaine D. Lorenzen,^{5,6,7*} Matteo Fumagalli,^{8,9} Bo Li,^{1,2,3} Kelley Harris,⁶ ZJun Xiong,¹ Long Zhou,¹ Thorfinn Sand Kornelussen,⁴ Mehret Somel,^{4,10} Courtney Babbitt,^{4,11,12} Greg Wray,^{4,12} Jianwen Li,¹ Weiming He,^{1,13} Zhuo Wang,¹ Wenjing Fu,¹ Xueyan Xiang,^{1,4} Claire C. Morgan,⁹ Aoife Doherty,¹⁰ Mary J. O'Connell,⁶ James O. McInerney,¹⁴ Erik W. Born,¹⁵ Love Dalén,¹⁶ Rune Dietz,¹⁷ Ludovic Orlando,⁴ Christian Sonne,¹⁸ Guojie Zhang,^{1,14} Rasmus Nielsen,^{1,3,19,20*} Eske Willerslev,^{4,21} and Jun Wang^{1,3,17,22,23,24}

¹BGI-Shenzhen, Shenzhen 518083, China

²School of Bioscience and Biotechnology, South China University of Technology, Guangzhou 510641, China

³Department of Integrative Biology, 3050 Valley Life Sciences Building, University of California, Berkeley, CA 94720, USA

⁴Centre for GeoGenetics, Natural History Museum, University of Copenhagen, Øster Voldgade 5-7, 1350 Copenhagen K, Denmark

⁵Department of Mathematics, 970 Evans Hall, University of California, Berkeley, CA 94720, USA

⁶Department of Biology, 124 Science Drive, Duke Box # 90338, Duke University, Durham, NC 27708, USA

⁷Institute for Genome Sciences & Policy, 101 Science Drive, DUMC Box 3362, Duke University, Durham, NC 27708, USA

⁸College of Life Sciences, Sichuan University, Chengdu 610064, China

⁹Bioinformatics and Molecular Evolution Group, School of Biotechnology, Dublin City University, Glasnevin, Dublin 8, Ireland

¹⁰Bioinformatics and Molecular Evolution Unit, Department of Biology, National University of Ireland, Maynooth, Co. Kildare, Ireland

¹¹Greenland Institute of Natural Resources, c/o Government of Greenland Representation in Denmark, Strandgade 91, 3. Floor, PO Box 2151, 1016 Copenhagen K, Denmark

¹²Department of Bioinformatics and Genetics, Swedish Museum of Natural History, PO Box 50007, 10-005, Stockholm, Sweden

¹³Department of Bioscience, Arctic Research Centre, Aarhus University, Frederiksborgvej 399, PO Box 358, 4000 Roskilde, Denmark

¹⁴Centre for Social Evolution, Department of Biology, University of Copenhagen, Universitetsparken 15, 2100 Copenhagen, Denmark

¹⁵Department of Statistics, 367 Evans Hall, University of California, Berkeley, CA 94720, USA

¹⁶Department of Biology, University of Copenhagen, Ole Mazowsz Vej 5, 2200 Copenhagen Ø, Denmark

¹⁷Princess Al Jawahra Center of Excellence in the Research of Hereditary Disorders, King Abdulaziz University, Jeddah 21589, Saudi Arabia

¹⁸Macao University of Science and Technology, Avenida Wai Long, Taipa, Macau 999078, China

¹⁹Department of Medicine, University of Hong Kong, Sassoon Road, Pokfulam, Hong Kong

²⁰Co-first authors

²¹Present address: Middle East Technical University, Department of Biological Sciences, 06800, Ankara, Turkey

²²Present address: Department of Biology, 611 North Pleasant St, University of Massachusetts Amherst, Amherst, MA, 01003, USA

²³Correspondence: rasmus_nielsen@berkeley.edu (R.N.), willerslev@nrm.ku.dk (E.W.), wang@genomics.org.cn (J.W.)

²⁴<http://dx.doi.org/10.1016/j.cell.2014.03.054>

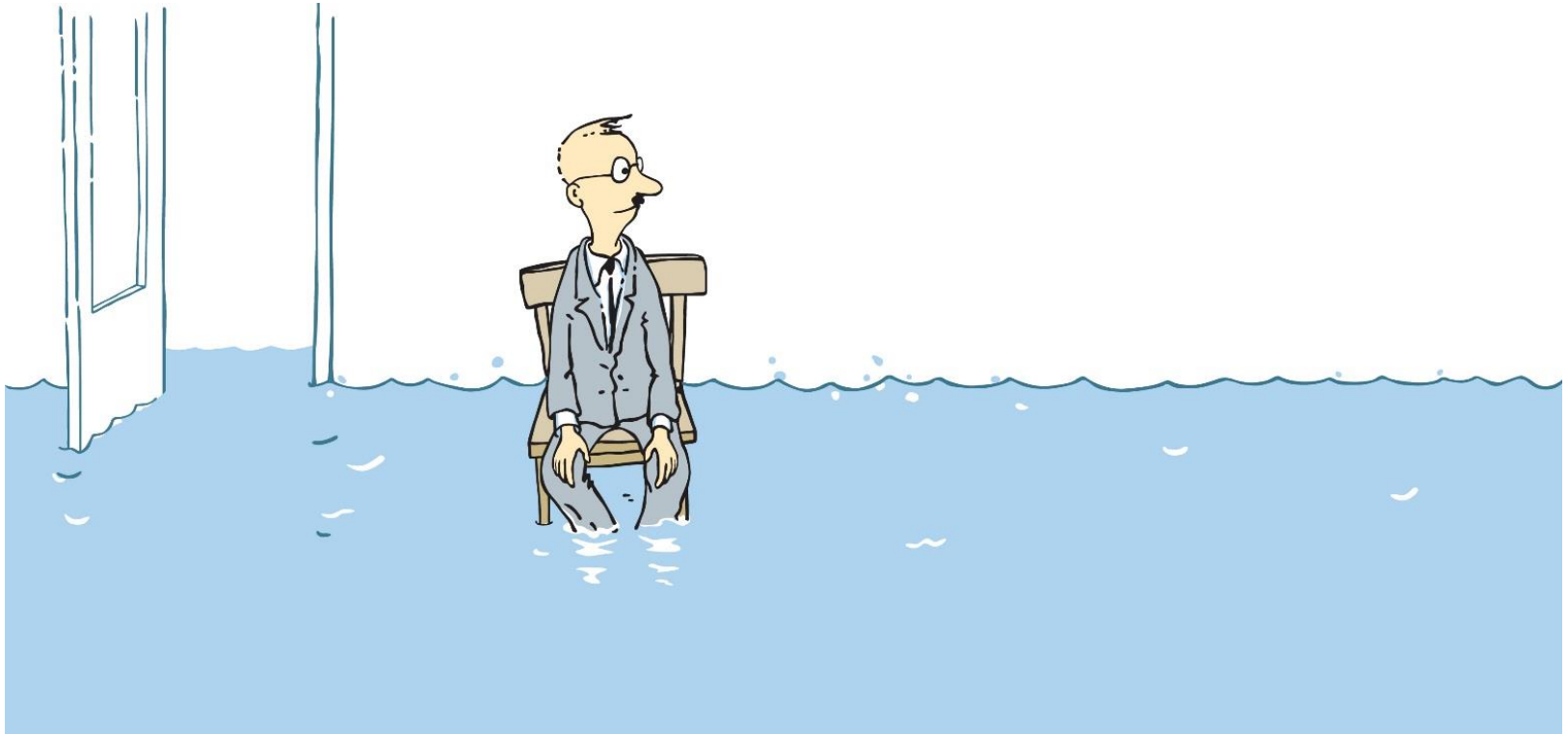
SUMMARY

Polar bears are uniquely adapted to life in the High Arctic and have undergone drastic physiological changes in response to Arctic climates and a hyper-lipid diet of primarily marine mammal prey. We analyzed 88 complete genomes of polar bear and brown bear using population genomic modeling and show that the species diverged only 479–343 thousand years BP. We find that genes on the polar bear lineage have been under stronger positive selection than in brown bears; nine of the top 16 genes under strong positive selection are associated with cardiomyopathy and vascular disease, implying important reorganization of the cardiovascular system. One of the genes showing the strongest evi-

how polar bears are able to cope with life-long elevated LDL levels that are associated with high risk of heart disease in humans.

INTRODUCTION

The polar bear (*Ursus maritimus*) is uniquely adapted to the extreme conditions of life in the High Arctic and spends most of its life out on the sea ice. In cold Arctic climates, energy is in high demand. Lipids are the predominant energy source and the polar bear has a lipid rich diet throughout life. Young nurse on milk containing ~27% fat (Hedberg et al., 2011) and adults feed on a marine mammal diet, primarily consisting of seals and their blubber (Thiassen et al., 2008). Polar bears have substantial adipose deposits under the skin and around organs, which can comprise up to 50% of the body weight of an individ-



The water is rising quickly. Should the man wait for delivery of a complex pump that's on a ten-year backorder from the hardware store? Or should he *kick a hole in the wall* to let the water drain out?

Darwinian
evolution
at work



**Key
Concepts
from
*Darwin
Devolves***

The First Rule of
Adaptive Evolution

The Principle of
Comparative Difficulty

The Family Line

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The Principle of Comparative Difficulty

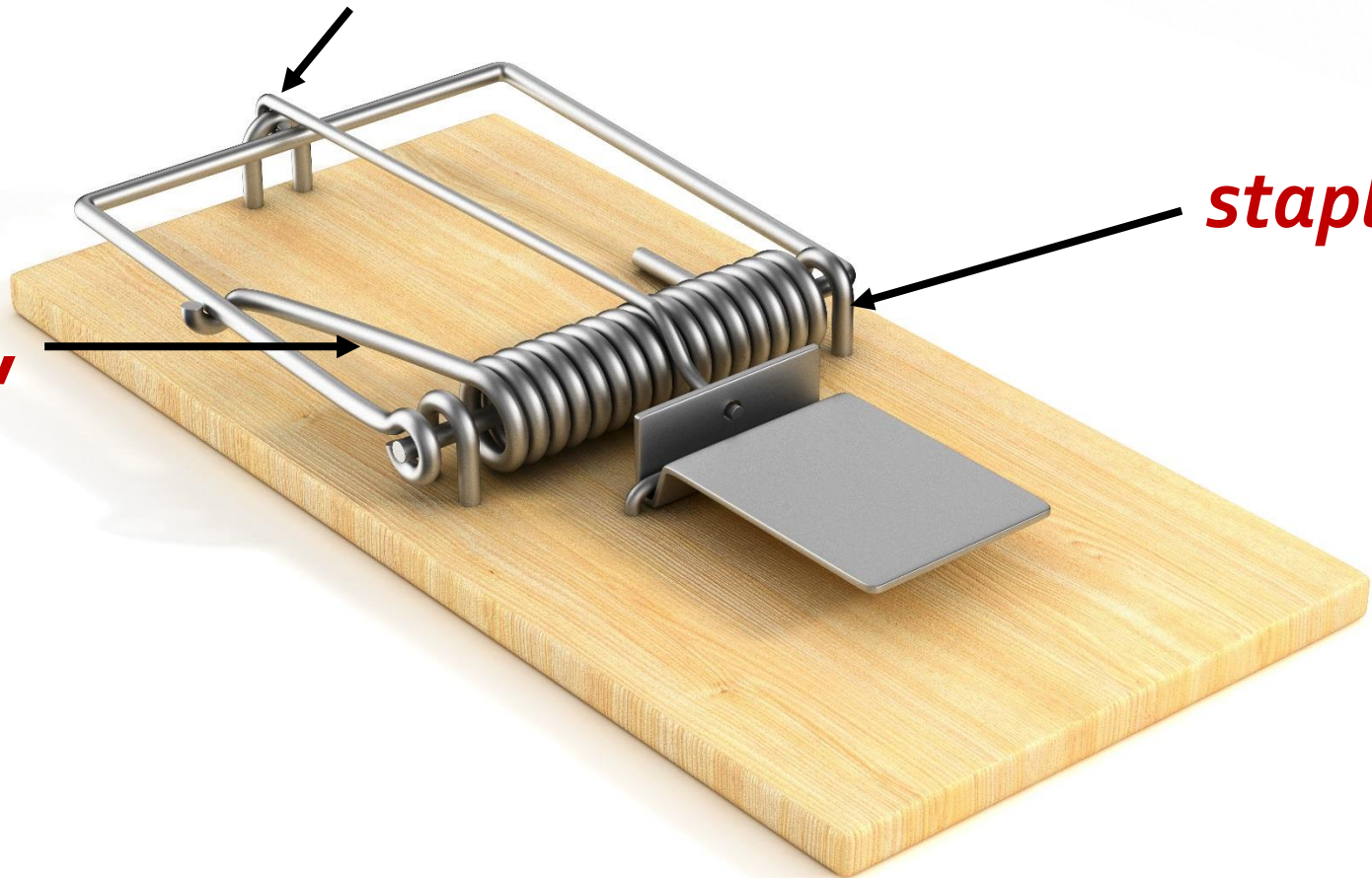
*If a task that requires **less** effort
is too difficult to achieve,
then a task that requires much
more effort necessarily is too.*

A common mechanical mousetrap needs multiple pieces *that are themselves complex*

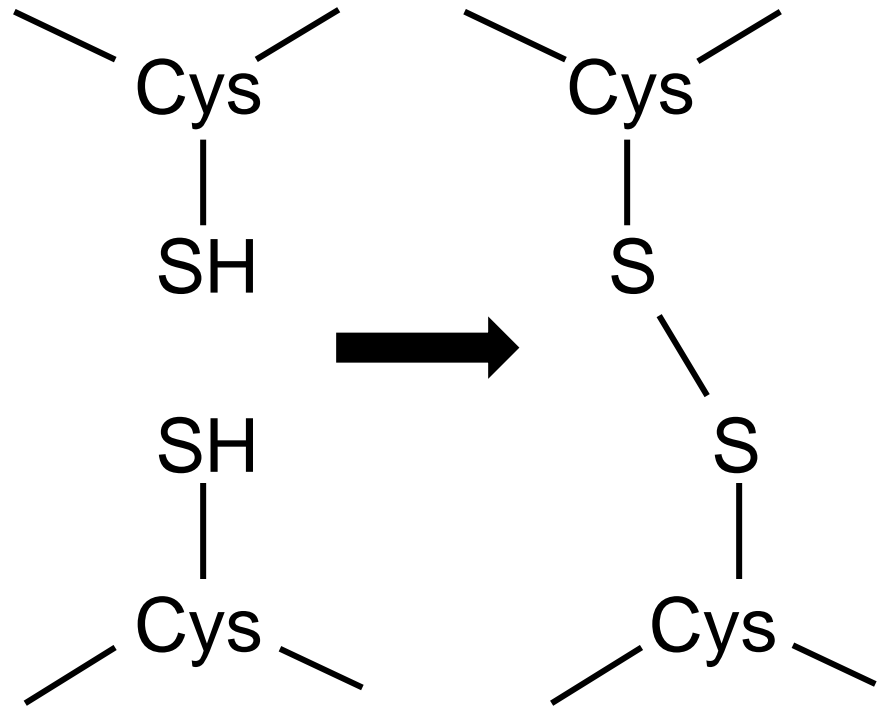
holding
bar *curl*

spring
length,
angle

staples



mini Irreducible Complexity (mIC):



Behe, M.J., Snoke, D.W., 2004. Simulating evolution by gene duplication of protein features that require multiple amino acid residues. *Protein Sci* 13: 2651-2664.

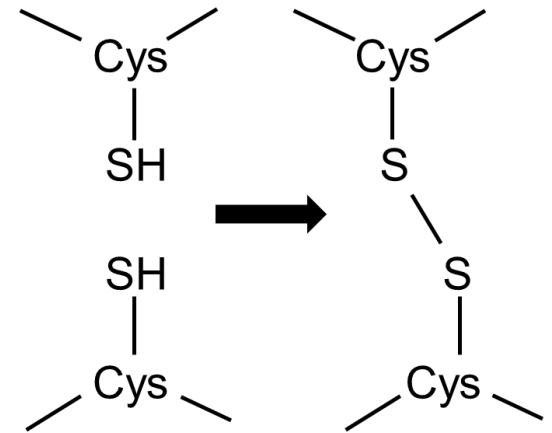
“We conclude that, in general, to be fixed in 10^8 generations, the production of novel protein features that require the participation of two or more amino acid residues simply by multiple point mutations in duplicated genes would entail population sizes of no less than 10^9 .”



Darwinism is claimed to explain:

- cells
- the genetic code
- molecular machines
- genetic networks
- phyla
- literature, music
- politics, the law
- love, the universe
- *even mind itself*

- *It just has trouble explaining a disulfide bond*



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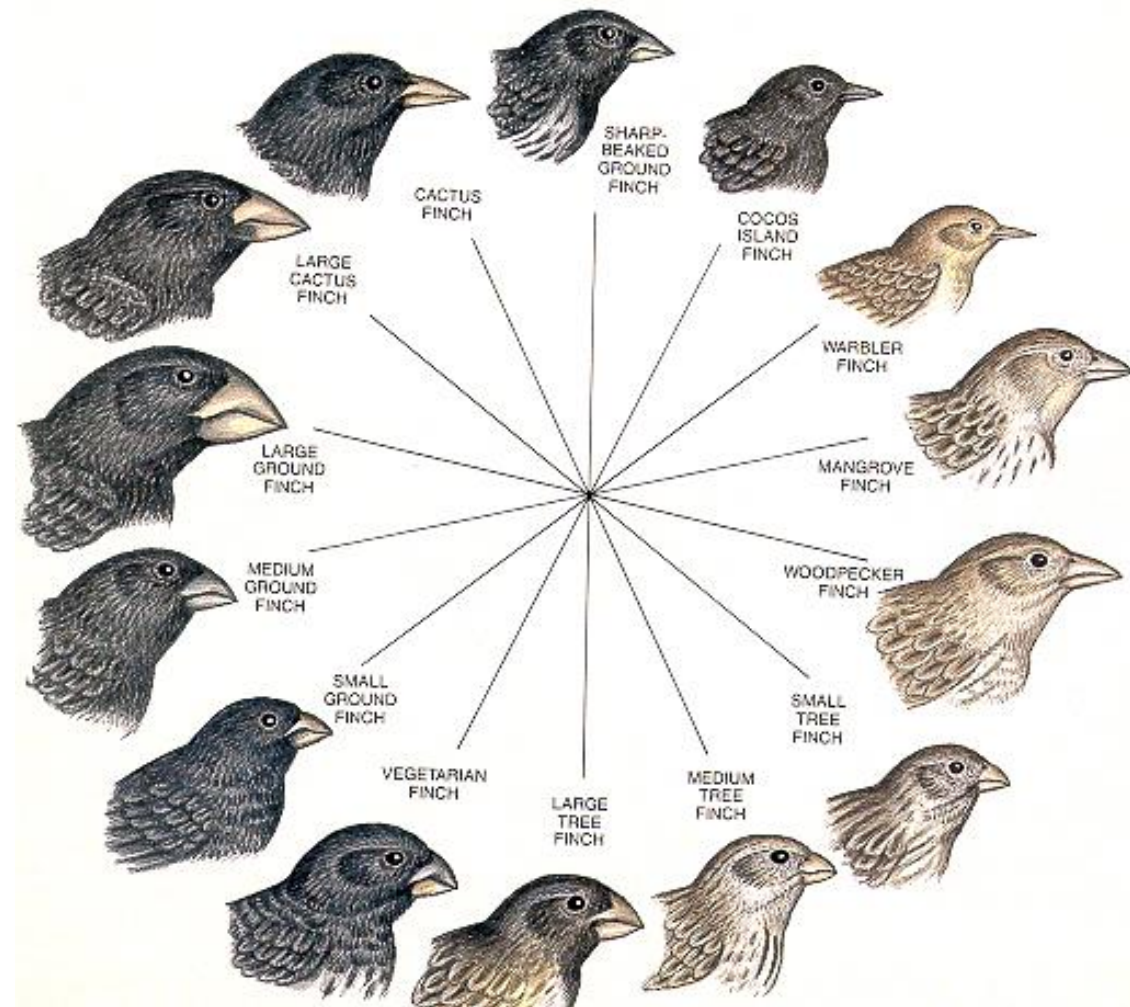
The Family Line

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The Family Line



Galápagos finch species

Classification of Galápagos finches and their ancestor

Level	Ancestor	Descendant
Domain	Eukaryota	Eukaryota
Kingdom	Animalia	Animalia
Phylum	Chordata	Chordata
Class	Aves	Aves
Order	Passeriformes	Passeriformes
Family	Thraupidae	Thraupidae
Genus	unknown	<i>Geospiza; Camarhynchus; Certhidea; Pinaroloxias</i>
Species	unknown	Various

Two million years of relentless evolution

\$213,754.36



\$213,754.83

Species of Cichlid fish

Lake Tanganyika



Julidochromis ornatus



Tropheus brichardi



Bathybates ferox



Cyphotilapia frontosa



Loboichilotes labiatus

Lake Malawi



Melanochromis auratus



Psudotropheus microstoma



Ramphochromis longiceps



Cyrtocara moorei



Placidochromis milomo

Classification of African great lake cichlids and their ancestor

Level	Ancestor	Descendant
Domain	Eukaryota	Eukaryota
Kingdom	Animalia	Animalia
Phylum	Chordata	Chordata
Class	Actinopterygii	Actinopterygii
Order	Perciformes	Perciformes
Family	Cichlidae	Cichlidae
Genus	unknown	various
Species	unknown	various

New classifications produced by luxuriantly evolving groups

group	species	genera	families	higher
finches	14	4	0	0
cichlids	~1500	~75	0	0
anoles	~300	3	0	0
honeycreepers	55	24	0	0
fruit flies	~1000	2	0	0
beetles	239	1	0	0
silverswords	50	3	0	0
lobelias	126	6	0	0

The Family Line:

Darwinian evolution is *self-limiting*

Three factors *restrict* it at the *highest* levels of biology
by *promoting* it at the *lowest* levels:

Random mutation

Natural selection

Irreducible complexity

The interaction
of intelligent &
unintelligent
evolutionary
processes

The *First Rule of Adaptive Evolution* allows organisms to quickly adapt to their environment by devolutionary processes.

Undirected evolutionary change rapidly bogs down before the *Family Line*.

Prediction: Unique information, inaccessible to unguided processes, is required at the level of family and higher.

The recognizable depth of design in animals as of **AD 2019**
The limit of Darwinian evolution now seems to
be at the biological level of *family*.

Families in the mammalian Order Carnivora





lehigh.edu/.../behe.html

discovery.org

michaelbehe.com

My responses
to critics can
be found at:



A MOUSETRAP FOR
DARWIN

2020

MICHAEL J. BEHE
ANSWERS HIS CRITICS

